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Rearing and maintaining Austrian PCN populations at AGES

Ines Gabl, Hermann Hausdorf

Department for Healthy Plants in Arable Crops and Horticulture

Occurrence of PCN in Austria

First findings of PCN in Austria







- start of living PCN collection in 2013
- cysts which were multiplied in the bioassay (standard procedure for soil testing at AGES until 2011)
- cysts from infested fields which were collected during re-sampling
 - cyst amount per 250 ml soil:
 approximately 1 5 cysts with living cyst content



rearing/multiplication - maintaining



- rearing/multiplication
 - as many cysts as possible for inoculum use for experiments
 - only used for cysts which are multiplied in the bioassay

- maintaining
 - keep the population alive with as less effort as possible
 - only used for extracted cysts from fields (1 5 living cysts per 250 ml soil)

rearing/multiplication - maintaining

- circumstances are the same
- test procedures for rearing/multiplication and maintaining are the same
 - pot test in greenhouse with susceptible potato varieties
 - controlled climate conditions
 - controlled water supply
- preparing inoculum density
- cyst extraction with MEKU soil sample extractor
- ethanol method
- investigation of produced cyst amount and cyst content
- storage
- difference in pot size (used soil amount) and inoculum density



Rearing/multiplication - material & method

- greenhouse pot test starts in February
- susceptible variety "Desiree"/ locally used susceptible variety "Hermes"
- tuber size 35/45 mm
- 2 litre pots
- strong paper liner is used to prevent roots from growing out
- inoculum density: 5 eggs and larvae/ml soil
- cyst inoculum on basis of the mean cyst content on a small batch of cysts (cysts from bioassay samples)



rearing/multiplication - material & method



- pot is filled with 1000 ml soil
- prepared cysts are stirred in the soil with a glass rod
- the tuber is planted with one sprout and covered with the remaining 500 ml soil
- 🦰 total soil amount: 1500 ml

rearing/multiplication - greenhouse conditions



- temperature regime:
 - 17,5°C-19,5°C (room temperature)
 - temperature measured in pots
- light regime: 16 h (D), 8 h (N)
 - → climate is monitored continously
- water regime:
 - before sprouts emerge water apply is done manually (2-3 x/week)
 - after sprouting automatic water supply (2-3x/week)

rearing/multiplication - material & method

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rearing/multiplication - material & method

- plants are tied to a stick to prevent them from toppling down
- extra sprouds emerging from tubers are removed
- only one stem is allowed to grow



- pest monitoring with yellow and blue sticky card traps
- pest control with use of benificial insects
 - Encarsia formosa greenhouse whitefly
 - · Chrysopa carnea aphids
 - Neoseiulus cucumeris thrips



rearing/multiplication - material & method



- after approximately 12 weeks potato plants wilt and die
- mater supply is stopped
- above ground parts are removed
- soil dries in pots for at least two weeks (greenhouse)
- pots are stored in a climate chamber at 8°C, until elutriation of cysts

MEKU – soil sample extractor

- 1500 ml soil is divided in 6 portions
- the extracted material is washed in a porcelain bowl
- floating material is transferred into a funnel with filter paper
- floating material with cysts dries at room temperature











Ethanol method

- dry floating material with cysts is transferred into a 500 ml Erlenmeyer flask containing a rubber stopper on a wire
- ethanol is added
- the ethanol with debris is stirred well and wait for a few minutes
 - > cysts float on the surface
- pour flouting material onto a filter paper in a glass funnel
- remaining organic matter and cysts dry









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storage

- cysts are rolled on a filter paper and counted
- cysts are stored dry in labelled glass tubes at 4°C in laboratory refrigerator with constant temperature regime









Maintaining on the host plant

- equal test procedure as for rearing and multiplication
- only difference:
 - · pot size: 500 ml
 - soil amount: 250 ml
 - inoculum density: after extraction of soil samples 1 - 5 cysts with living content
 - but not more than 8 viable cysts per 250 ml soil





Populations and data

- 🦰 G. rostochiensis populations
- G. pallida populations
- mixed G. rostochiensis and G. pallida populations
- available data: federal state, field locality, field name, size of field, collecting year, collector, year of multiplication and storage
- populations are different concerning locality and region
- every five years action is taken to keep the population alive
- identity is confirmed at the beginning of maintaining, rearing/multiplication



Ines Gabl, He

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Department for Healthy Plants in Arable Crops and Horticulture

AGES – Austrian Agency for Health & Food Safety

Spargelfeldstraße 191 A-1220 Vienna T +43 (0)50 555-33315

ines.gabl@ages.at, hermann.hausdorf@ages.at www.ages.at