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Agroscope

# Risk-based sampling of woody plants for planting

# Case Study "Xylella fastidiosa"

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www.agroscope.ch I good food, healthy environment



Boscia, D. (2013). Symptoms of quick decline



Boscia, D. (n.d.). Symptoms of quick decline

#### October 2013



Boscia, D. (n.d.). Symptoms of quick decline

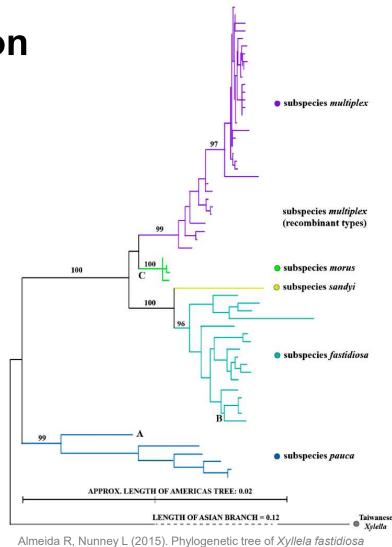
April 2013



Boscia, D. (n.d.). Symptoms of quick decline

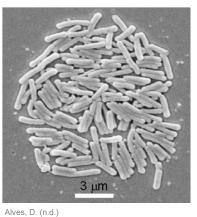
### Xylella fastidiosa - Introduction

- 3 major subspecies
- Over 560 host plant species
  - But specific to different isolates
- Median yield losses (EFSA, 2019)
  - Olives ≥ 30y : 69.1%
  - Citrus spp. : 13.9%
  - Grapes : 2.1%
- Potential economic impact : 5.5 B € /y in
  - EU in the future (EFSA, 2019)



#### Xylella fastidiosa - Interactions U

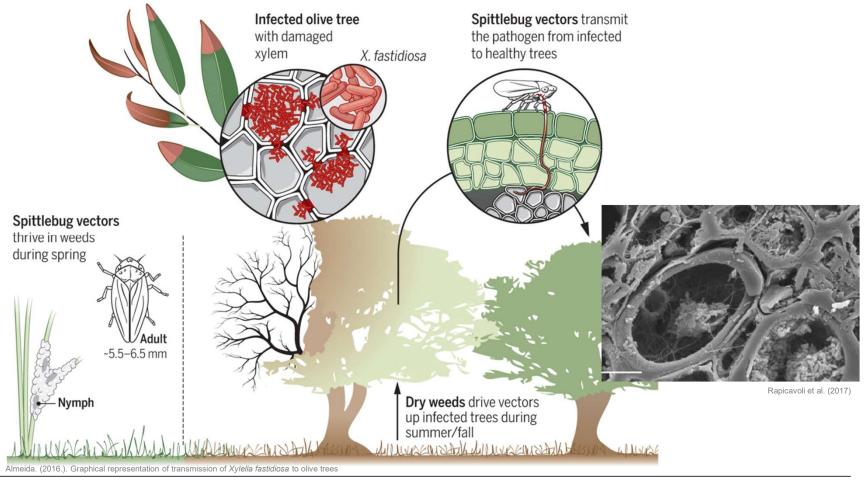






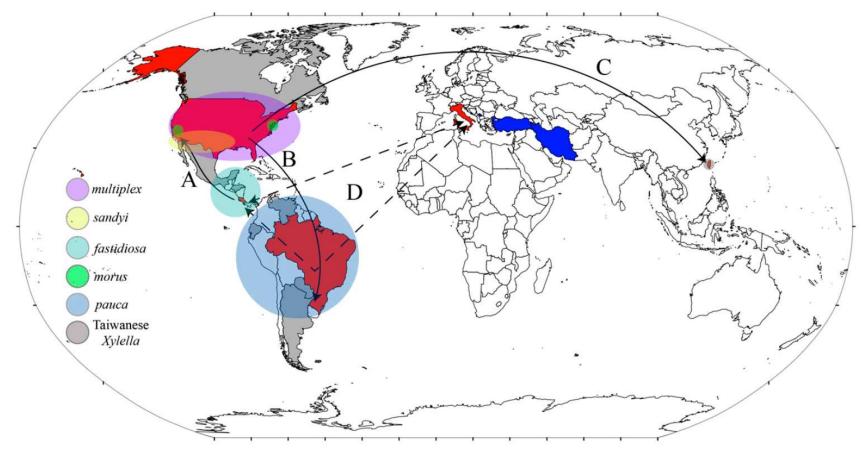
Klejdysz, T. (n.d.)

## Xylella fastidiosa - Transmission



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### **V** Xylella fastidiosa - Distribution



Almeida R, Nunney L (2015). World map illustrating the proposed endemic distribution of Xylella fastidiosa subspecies.

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## **V** Xylella fastidiosa – control measures

- Remove and burn infested plants
- Antibacterial treatment
- Insecticides (pyrethroid)
- R&D
  - Kaolin, Harpine, Beauveria bassiana
  - Resistant/ tolerant cultivars
- Prevent movement of infected plants
- Surveillance

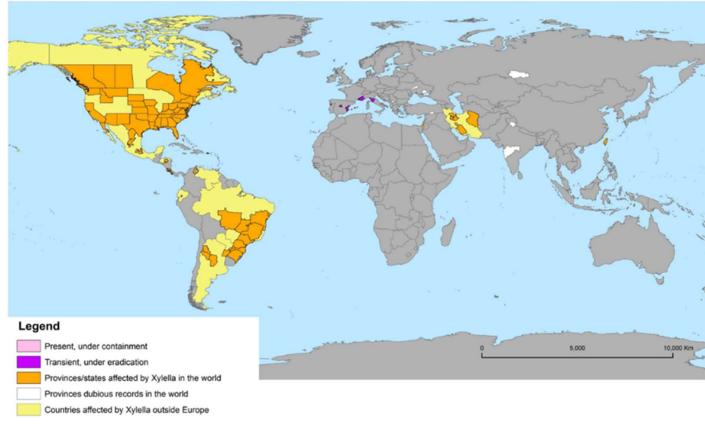
#### Risk estimation – Plant species

- Main host plant species as per 2015/789/EU:
  - Coffea spp.
  - Lavandula dentata L.
  - Nerium oleander L.
  - Olea europaea L.
  - Polygala myrtifolia L.
  - Prunus dulcis

- Other host plant species
- EUR-Lex 32020R1201 EN -

EUR-Lex (europa.eu)

#### Risk estimation – Country of origin



World distribution of Xylella fastidiosa (modified from EFSA PLH Panel, 2018)

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#### Pest survey card on Xylella fastidiosa (arcgis.com)



<u>ph\_biosec\_legis\_list-</u> <u>demarcated-union-</u> <u>territory\_en.pdf (europa.eu)</u>



# Risk estimation – Country of origin

- Three scenarios for importing plants
  - Country free of X. fastidiosa
    - Sampling only if symptoms are observed
  - Region free of X. fastidiosa in a country affected by X. fastidiosa
    - Routine samples
  - Region affected by X. fastidiosa
    - Import prohibited

# Hygenic measures

How is Xylella fastidiosa spread naturally?

Vector

- Routine sample  $\rightarrow$  basic hygienic measures
  - Avoid touching plants
  - Disinfection: tools, shoes, hands, sample bag
- Suspicion  $\rightarrow$  additional hygienic measures
  - E.g. disposable suit
  - Extended disinfection

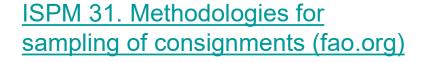


# Sampling of plants

Number of plants to be sampled

- Confidence level
- Level of detection
  - $\rightarrow$  both determined by NPPO
- Number of units in lot

Number of units in lot	P = 95% (confidence level) % level of detection × efficacy of detection				P = 99% (confidence level) % level of detection × efficacy of detection					
10.000	5	2	1	0.5	0.1	5	2	1	0.5	0.1
25	24*	-	-	-	-	25*	-	-	-	-
50	39*	48	-	-	-	45*	50	-	-	-
100	45	78	95	-	-	59	90	99	-	-
200	51	105	155	190	-	73	136	180	198	-
300	54	117	189	285*	-	78	160	235	297*	-





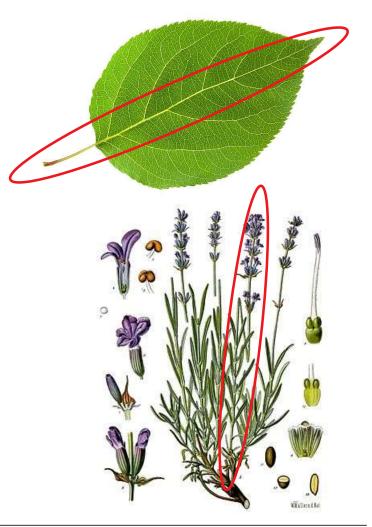
# Sampling of plants

What type of plant tissue is to be sampled?

Xylem containing plant tissue

Sampling adapted according to

- Period of active growth
- Host plant
- What the lab needs



# Sampling of plants

Presence of symptoms: Suspicion

- Individual plants
- Pool only similar symptoms
- Branches with 10–25 leaves
- Transitional tissue

Absence of symptoms: Routine sample

- Representative sample
- 4–10 branches for individual plants

#### DP 25: Xylella fastidiosa (fao.org)

Table 1. Number of leaves (including their petioles) to be used and approximate weight of the laboratory sample Data from EPPO (2018b).

Type of sample	Host plants and type of tissue	Minimum number of leaves per laboratory sample	Approximate weight of laboratory sample	
Sample from individual plant with leaves	Petioles or midribs, or both, of leaves of large size (e.g. from Coffea spp., Ficus spp., Vitis spp., Nerium spp.)	5	0.5–1 g	
	Petioles or midribs, or both, of leaves of small size (e.g. Polygala myrtifolia and Olea spp.)	25	0.5–1 g	
	Plant species without petioles or with small petiole and midrib	25	0.5–1 g	
Dormant plant or cuttings Xylem tissue		n/a†	0.5-1 g	
Composite sample from several coffee plants from a single lot with leaves	Samples of asymptomatic plants (e.g. collected from imported consignments or nursery monitoring)	100–200	10–50 g	

#### PM3/081(3) Inspection of consignments for Xylella fastidiosa (eppo.int)

TABLE 1 Guidance on sampling for lots of plants for selected species and tissue to be recovered when testing samples composed of large amounts of tissue\* (e.g. composite samples from consignment/ places of production of plants for planting)

Host	Minimum number of leaves/twigs/stems to be collected per plant	Number of plants that can be pooled		
Olea europaea <sup>b</sup>	4 (leaves)	Up to 225		
Nerium oleander	2 (leaves)	Up to 100		
Herbaceous plantlets	l (plantlet)	Up to 200		
Polygala myrtifolia <sup>e</sup>	2 (twigs)	Up to 125		
Lavandula spp.4	2 (stems)	Up to 100		
Prunus dulcis/P. avium	2 (twigs)	Up to 100		
Coffea spp.	2 (leaves)	Up to 50		
Helichrysum italicum	2 (stems)	Up to 50		

## Transport and shipment

- Threefold packaging: 2 layers of plastic + stable container
- Unique label

20230427\_12345\_1 Lavendula angustifolia Routine sample XYLEFA

- Cooled transport (4–15 °C)
- As fast as possible



#### Practical exercise

You are called by the customs at Zurich airport for the following situations. What are the procedures to be followed in each situation? Confidence level = 95%, detection level = 1%

State (country) of origin	Plant species	Lot size	Risk based inspection method	Sample size	Minimum number of leaves/twigs/stems to be collected per plant	Number of plants that can be pooled
Oregon (USA)	Prunus dulcis	500				
California (USA)	Prunus dulcis	500				
Fès-Meknès (MA)	Olea europea	50				
Bahia (BR)	Theobroma cocoa	100				
Western Cape (SA)	Polygale myrtifolia	1500				







#### Thank you for your attention

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